

Curriculum Vitae

Kathleen Mulvaney

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EDUCATION

UNIVERSITY OF NORTH CAROLINA CHAPEL HILL
PhD in Cell Biology
Dissertation: Proteomic dissection of KEAP1/NRF2 signaling identifies novel pathway interactors

CHAPEL HILL, NC
December 2016

UNIVERSITY OF ROCHESTER
Bachelors of Science in Biology
Concentration in Molecular Genetics with Distinction in Research

ROCHESTER, NY
May 2010

POSITIONS

FRALIN BIOMEDICAL RESEARCH INSTITUTE OF VIRGINIA TECH
Assistant Professor of Biomedical Sciences and Pathobiology
Assistant Professor of Biomedical Engineering

WASHINGTON, DC
October 2022-Present

- Research focus on targeting enzyme-substrate interactions for therapeutic development in cancer
- Therapeutically targeting PRMT5 in CDKN2A/MTAP null solid cancers

CHILDREN'S NATIONAL HOSPITAL
Affiliate Member, Cancer and Immunology Research Center

WASHINGTON, DC
October 2022-Present

- Research focus on targeting enzyme-substrate interactions for therapeutic development in cancer

ATRIUM WAKE FOREST BAPTIST COMP. CANCER CNTR
Member

WAKE FOREST, NC
February 2023-Present

- Therapeutically targeting PRMT5 in CDKN2A/MTAP null pediatric and adult cancer

THE BROAD INSTITUTE; DANA FARBER CANCER INST.
HARVARD UNIVERSITY

CAMBRIDGE, MA

Postdoctoral Fellow with Dr. William (Bill) Sellers

June 2017-September 2022

- Research focus on targeting protein-protein interactions for therapeutic development in cancer
- Therapeutically targeting PRMT5 on a novel binding interface in pancreatic cancer
- Led a productive team of 2-3 research associates funded by NIH and Deerfield Management support
- Collaborated with the Center for Development of Therapeutics to complete drug discovery project
- Culminated in two research publications and one patent

UNIVERSITY OF NORTH CAROLINA CHAPEL HILL
Graduate Student with Dr. Ben Major

CHAPEL HILL, NC
Spring 2011-June 2017

- Research focus on the KEAP1-NRF2 signaling pathway in cancer
- Described a function for KEAP1 in cell cycle regulation through ubiquitylation of MCM3
- Proteomic screening identified novel NRF2 interacting proteins and transcriptional regulators
- Culminated in successful defense of PhD, four research papers, and a review article

- Research Focus on Diffuse Large B-Cell Lymphoma and expression of MHC Class II and CIITA with the underlying goal of identifying the association between expression levels and patients' immunological responses to DLBCL tumors
- Culminated in writing and successful defense of senior thesis and two co-authored manuscripts

PUBLICATIONS

1. **Kathleen Mulvaney**. Early clinical success of MTA-cooperative PRMT5 inhibitors for the treatment of CDKN2A/MTAP deleted cancers. *Cancer Discovery*. **13**, 2310–2312 (2023). PMID: 37909092. Invited submission.
2. **Mulvaney, KM**; Blomquist, C; Acharya, N; Li, R; Ranaghan, MJ; O'Keefe, M; Rodriguez, DJ; Young, MJ; Kesar, D; Pal, D; Stokes, M; Nelson, AJ; Jain, SS; Yang, A; Mullin-Bernstein, Z; Columbus, J; Bozal, FK; Skepner, A; Raymond, D; ... Sellers, WR. Molecular basis for substrate recruitment to the PRMT5 methylosome. *Molecular Cell*. 2021, 81(17)17, P3481-3495.
3. McKinney, DC; McMillan, BJ; Ranaghan, MJ; Morocco, JA; Brousseau, M; Mullin-Bernstein, Z; O'Keefe, M; McCarren, P; Mesleh, M; **Mulvaney, KM**; Robinson, F; Singh, R; Bajrami, B; Wagner, FF; Hilgraf, R; Drysdale, MJ; Campbell, AJ; Skepner, A; Timm, DE; Porter, D; Kaushik, VK; Sellers, WR; Ianari, A. Discovery of a First-in-Class Inhibitor of the PRMT5–Substrate Adaptor Interaction. *Journal of Medicinal Chemistry*. 2021, 64 (15), 11148-11168.
4. Song, S; Nguyen, V; Schrank, T; **Mulvaney, KM**; Walter, V.; Wei, D.; Orvis, T.; Desai, N.; Zhang, J.; Hayes, DN; Zheng, Y.; Major, MB; Weissman, B. Loss of SWI/SNF chromatin remodeling alters NRF2 signaling in non-small cell lung carcinoma. *Mol Cancer Res*. 2020, Dec; 18(12):1777-1788.
5. Bowman BM; Montgomery SA Schrank TP, Simon JM, Ptacek TS, Tamir TY, **Mulvaney KM**, Weir SJ, Nguyen TT, Murphy RM, Makowski L, Hayes DN, Chen XL, Randell SH, Weissman BE, Major MB. A conditional mouse expressing an activating mutation in NRF2 displays hyperplasia of the upper gastrointestinal tract and decreased white adipose tissue. *J Pathol*. 2020 Oct;252(2):125-137.
6. **Mulvaney, KM**; Matson, J.; Siesser, P.; Tamir, TY; Goldfarb, D.; Jacobs, T.; Cloer, EW; Cook, JG; Major, MB. Identification and Characterization of MCM3 as a KEAP1 Substrate. *Journal of Biological Chemistry*, 2016 Nov 4; 291(45): 23719–23733.
7. Tamir, TY*; **Mulvaney, KM***; Major, MB. Dissecting the Keap1/Nrf2 pathway through proteomics. *Current Opinion in Toxicology*, (*equal contribution) 2016, 1:118-24.
8. Guntas, G; Lewis, S; **Mulvaney, KM**; Cloer, E; Tripathy, A; Lane, T; Major, MB; Kuhlman, B. Engineering a genetically encoded competitive inhibitor of the KEAP1-NRF2 interaction via structure- based design and phage display. *Protein Eng Des Sel*. 2016 Jan;29(1):1-9.
9. Hast, BE; Goldfarb D*; **Mulvaney KM***; Hast MA; Siesser PF; Yan F; Hayes DN; Major MB. Proteomic analysis of ubiquitin ligase KEAP1 reveals associated proteins that inhibit NRF2 ubiquitination. *Cancer Research*. 2013 1;73(7):2199-210. (*equal contribution)
10. Cycon, KA; **Mulvaney, KM**; Rimsza, LM; Persky, D.; Murphy, SP. Histone deacetylases (HDACs) contribute to the silencing of CIITA expression in DB diffuse large B cell lymphoma. *Immunology*. 2013 Oct; 140(2): 259–272.
11. Bushway, M.; Cycon, KA; **Mulvaney, KM**; Murphy, SP. Coordinate loss of MHC class II expression in the diffuse large B cell lymphoma cell line OCI-Ly2 is due to a novel mutation in RFX-AP. *Immunogenetics*. 2010, 62(2):109-16.

PATENTS

1. Arthur Campbell, Martin Drysdale, Robert Hilgraf, Alessandra Ianari, Patrick McCarren, David

McKinney, Brian McMillan, **Kathleen Mulvaney**, Dale Porter, William Sellers, Ritu Singh, Florence Wagner. Co-inventor, PCT Application Serial No. PCT/US2021/045016, filed on August 6, 2021 and entitled, "Substrate Adaptor Inhibitors of PRMT5 and Uses Thereof". (inventors listed alphabetically)

AWARDS AND FUNDING

- NIH R35 MIRA Grant Award: 1R35GM154987-01 (2024-2029)
- Children’s Cancer Foundation Grant (2024)
- Focused Ultrasound Foundation Pilot Grant (2024)
- Seale Innovation Pilot Grant (2023)
- NIH F32 Ruth L. Kirschstein Postdoctoral Individual National Research Service Award (2018-2021)
- NIH LRP Award (2019-2021)
- Sigma Xi Research Society Graduate Student Travel Award (2015)
- National Science Foundation Graduate Research Fellowship Honorable Mention (2011, 2012)
- Degree with Distinction in Research: an honor awarded by the University of Rochester (2009)
- National Science Foundation David T. Kearns Scholar (2008-2010)
- Take Five Scholar: Awarded tuition-free fifth year by the University of Rochester to study “Influences on Cognitive and Personality Development” (2009- 2010)
- Academic Competitiveness Grant awarded by New York State (2007- 2008)
- International Baccalaureate Scholar awarded as a four-year merit scholarship (2005- 2009)

PRESENTATIONS

NATIONAL MEETINGS

- FASEB Biological Methylation Conference** February 2023
- Presented “PRMT5 as a therapeutic target in CDKN2A/MTAP null cancer” invited talk
- FASEB Biological Methylation Conference** June 2022
- Presented “PRMT5 as a therapeutic target in CDKN2A/MTAP null cancer” talk
- AACR Annual Meeting** April 2022
- Presented “Resolving substrate recruitment to the PRMT5 methylosome” invited seminar
- AACR Annual Meeting** April 2016
- Presented “KEAP1 ubiquitinates MCM3 to regulate cell cycle in cancer” poster
- FASEB Ubiquitin Conference** June 2014
- Presented “KEAP1 ubiquitinates MCM3” poster to the ubiquitin-proteasome signaling community

DANA FARBER HARVARD CANCER CENTER

- Mass. General Hospital Postdoc Association Invited Seminar** February 2022
- Presented “Drugging Enzymes in Cancer” to a general audience, invited seminar
- Ludwig Meeting Invited Seminar** July 2021
- Presented “Resolving substrate recruitment to the PRMT5 methylosome” invited seminar
- Molecular and Cellular Oncology Retreat** June 2019
- Awarded Best Poster Presentation: “PRMT5-substrate adaptor interface as a novel therapeutic target in MTAP null tumors”

BROAD INSTITUTE

- Lunch and Learn** July 2019
- Co-led department-wide invited chalk talk: “Opportunities and challenges in following up cancer dependencies: lessons learned from PRMT5 and NXT1”

UNIVERSITY OF NORTH CAROLINA CHAPEL HILL

- University Research Day** March 2015

- Awarded Best Oral Presentation

November 2014

IMSD Symposium

- Awarded Best Oral Presentation

UNIVERSITY OF ROCHESTER

Office of Admissions

December 2008

- Invited to present regionally to prospective students on undergraduate research opportunities

TEACHING EXPERIENCE

VIRGINIA POLYTECHNIC INS. & STATE UNIVERSITY, BLACKSBURG, VA

Graduate Student Mentoring

Spring 2023

- Mentoring Virginia Tech TBMH students rotating in the lab in critical thinking and scientific research

Lecturer

Spring 2023

- Presented lecture on defining novel therapeutic targets in cancer in Graduate Cancer Focus Class

HARVARD UNIVERSITY, CAMBRIDGE, MA

BIOS E-30 Epigenetics & Gene Regulation

Teaching Assistant and Guest Lecturer

Fall 2020, Fall 2021

- Led weekly independent recitation and study sections, guided and graded the writing assignments
- Teaching 2 lectures per seminar on cancer epigenetics (2021)

BIOS E-16 Cell Biology

Teaching Assistant

Spring 2019; Spring 2021

- Led weekly independent recitation and study sections
- Guided and graded the writing assignments, problem sets, and exams

BROAD INSTITUTE, CAMBRIDGE, MA

Graduate Student Mentoring

September 2017-Present

- Mentoring Harvard BBS students rotating in the lab in critical thinking and scientific research

Research Associate Mentoring

June 2017-Present

- Mentoring and promoting the career development of three post-undergraduate researchers in the lab.

UNIVERSITY OF NORTH CAROLINA, CHAPEL HILL, NC

Science-at-Hand

October 2014-May 2017

- Give monthly scientific demonstrations and discuss careers in science with high school students.

Undergraduate Mentoring

January 2014- May 2017

- Mentoring two undergraduate researchers on research projects for credit in the Major Lab.

First Year Group

Fall 2014

- Peer mentor to first year students, advise on lab rotations and giving scientific presentations.

North Carolina DNA Day

April 2011; April 2013

- Taught high school students about pharmacogenomics and encouraged exploring careers in science.

LABORATORY TECHNIQUES

PROTEOMICS: Experience with purification of protein complexes for identification by mass

spectrometry (affinity, immunoprecipitation and BioID), isolation and detection of whole cell and targeted post-translational modifications (phosphorylation, ubiquitylation, methylation) by mass spectrometry, and analysis and visualization of mass spectrometry data (MaxQuant, Proteome Discoverer, Skyline, Spotlite, Cytoscape).

GENOMICS, SCREENING AND NEXT-GEN SEQUENCING: Experience with single and digenic CRISPr knockout screens, small molecule screening, RNA-seq, library generation, and Illumina sequencing.

MOLECULAR BIOLOGY & BIOCHEMISTRY: Experience with mammalian cell culture (primary and immortalized), RNA and DNA isolation, qPCR, RNA-seq, genetic manipulation including CRISPr, cloning and site-directed mutagenesis, flow cytometry, fluorescence activated cell sorting, chromatin immunoprecipitation, luciferase reporter assays, drug treatments and viability and proliferation assays, protein expression and purification, Western blotting, fixed and live-cell light and fluorescence microscopy, proximity ligation assays, mouse handling, MEF line derivation, subcutaneous xenografts, recombinant protein purification, size exclusion chromatography, and cellular and *in vitro* binding and ubiquitylation and methylation assays.

PROFESSIONAL MEMBERSHIPS

American Association for Cancer Research (AACR) (2016-present)

American Association for the Advancement of Science (AAAS) (2015-present)

JOURNAL REVIEWER

Molecular Cancer Research (2019-present)

PLOS One (2023-present)